

Instruction Manual

Ver. 09.26.18 For Research Use Only

Presto™ Mini RNA Yeast Kit

RBY004, RBYD004 (4 Preparation Sample Kit) RBY050, RBYD050 (50 Preparation Kit) RBY100, RBYD100 (100 Preparation Kit) RBY300, RBYD300 (300 Preparation Kit)

Advantages

Sample: a variety of yeast and other fungus species

Yield: up to 30 μ g of RNA (5 x 10⁷ Saccharomyces cerevisiae: 20 μ g)

Convenient: includes Sorbitol Buffer to reduce sample preparation time

Format: certified DNase and RNase-free spin columns

Time: within 20 minutes
Elution Volume: 50-100 μl

Kit Storage: dry at room temperature (15-25°C)

Table of Contents

Introduction	 2
·	
Troubleshooting	 5
_	



Introduction

The Presto™ Mini RNA Yeast Kit was designed for total RNA purification from yeast and a wide variety of other fungus species. Sorbitol Buffer is included with the kit to reduce sample preparation time and minimize hands on time. Detergents and chaotropic salt are used to lyse cells and inactivate RNase while RNA is bound by the glass fiber matrix of the RNA spin column. Once any contaminants have been removed, using the Wash Buffer (containing ethanol), the purified total RNA is eluted by RNase-free Water. High quality total RNA can be purified within 20 minutes and is ready for use in RT-PCR, Northern Blotting, Primer Extension, mRNA Selection and cDNA Synthesis.

Quality Control

The quality of the Presto[™] Mini RNA Yeast Kit is tested on a lot-to-lot basis by isolating RNA from *Saccharomyces cerevisiae* (5×10⁷) harvested by centrifugation at 5,000 x g for 10 minutes. A 5 µl aliquot of purified RNA from a 50 µl eluate is analyzed by electrophoresis on a 0.8% agarose gel.

Kit Components

Component	RBY004 RBYD004	RBY050 RBYD050	RBY100 RBYD100	RBY300 RBYD300
Sorbitol Buffer	4.5 ml	45 ml	45 ml x 2	225 ml
RB Buffer	2 ml	30 ml	60 ml	130 ml
DNase I ¹ (2U/μI) (RBYD004/050/100/300 Only)	20 µl	275 μΙ	550 µl	550 µl x 3
DNase I Reaction Buffer (RBYD004/050/100/300 Only)	200 μΙ	2.5 ml	5 ml	15 ml
W1 Buffer	2 ml	30 ml	50 ml	130 ml
Wash Buffer ² (Add Ethanol)	1.5 ml (6 ml)	25 ml (100 ml)	25 ml + 12.5 ml (100 ml) (50 ml)	50 ml x 2 (200 ml x 2)
RNase-free Water	1 ml	6 ml	15 ml	30 ml
RB Columns	4	50	100	300
2 ml Collection Tubes	8	100	200	600

¹DNase I is shipped at room temperature and should be stored at -20°C for extended periods after receiving the kit.

Steps to prevent RNase contamination

- 1. During operation, always wear a lab coat, disposable gloves, protective goggles and (anti-fog) procedure mask.
- Disposable plasticware and automatic pipettes should be sterile (RNase-free) and used only for RNA procedures.
- 3. Non-disposable glassware or plasticware should also be sterile (RNase-free).

²Add absolute ethanol (see the bottle label for volume) to Wash Buffer then mix by shaking for a few seconds. Check the box on the bottle. Be sure and close the bottle tightly after each use to avoid ethanol evaporation. The additional **Wash Buffer x 12.5 ml** is **only** included in **RBYD100**.





During the procedure, always wear a lab coat, disposable gloves, and protective goggles.

Quick Protocol Diagram



Cell lysis of yeast and other fungus species



RNA binding to membrane while contaminants remain suspended



Wash (removal of contaminants while RNA remains bound to membrane)



Elution of pure total RNA which is ready for subsequent reactions

Presto™ Mini RNA Yeast Kit Protocol

Please read the entire instruction manual prior to starting the Protocol Procedure.

IMPORTANT BEFORE USE!

Add absolute ethanol (see the bottle label for volume) to Wash Buffer then mix by shaking for a few seconds. Check the box on the bottle. Be sure and close the bottle tightly after each use to avoid ethanol evaporation

DNA Removal Options: For DNA-free RNA perform either option 1 (following RNA Binding) or option 2 (following RNA Elution).

Additional Requirements

lyticase or zymolase, absolute ethanol, ddH₂O (RNase-free and DNase-free) to prepare 70% ethanol, microcentrifuge tubes (RNase-free), pipette tips (RNase-free), ß-mercaptoethanol, EGTA (for DNA Digestion In Solution)



Yeast/Fungus Protocol Procedure

1. Sample Preparation

A. Yeast/Fungus on Agar Plate

Use an inoculating loop to transfer a small piece of yeast/fungus (up to 5 x 10⁷) from an agar plate to a 1.5 ml microcentrifuge tube containing 600 µl of Sorbitol Buffer.

B. Yeast/Fungus in Broth

Transfer yeast/fungus cells (up to 5 x 10^7) in broth to a 1.5 ml microcentrifuge tube. Centrifuge for 10 minutes at 5,000 x g then discard the supernatant. Re-suspend the cells in 600 μ l of Sorbitol Buffer.

Add **200 U of lyticase or zymolase** then mix well. Incubate at 30°C for 30 minutes. Centrifuge the mixture for 10 minutes at 2,000 x g to form a spheroplast pellet. Discard the supernatant.

2. Cell Lysis

Add 300 µl of RB Buffer and 3 µl ß-mercaptoethanol (or 6 µl of freshly prepared 2M Dithiothreitol in RNase Free Water) to the spheroplast pellet then vortex to mix. Incubate at room temperature for 5 minutes then centrifuge at 14-16,000 x g for 2 minutes. Transfer the supernatant to a new 1.5 ml microcentrifuge tube (RNase-free).

RNA Binding

Add 500 μ I of 70% ethanol to the lysate and pipette immediately. Place a RB Column in a 2 ml Collection Tube. Transfer 500 μ I of the mixture to the RB Column. Centrifuge at 14-16,000 x g for 1 minute then discard the flow-through. Transfer the remaining mixture to the same RB Column and centrifuge at 14-16,000 x g for 1 minute. Discard the flow-through and place the RB Column in a new 2 ml Collection Tube.

Optional Step 1: In Column DNase I Digestion

DNA contamination is significantly reduced following In Column DNase I Digestion. However, traces of residual DNA may be detected in very sensitive applications. In this situation, please perform Optional Step 2: DNA Digestion In Solution instead to efficiently remove trace amounts of DNA. Standard DNase buffers are incompatible with In Column DNase I Digestion and may affect RNA integrity and reduce yield.

- 1. Add 400 μ l of Wash Buffer (make sure ethanol was added) to the RB Column then centrifuge at 14-16,000 x g for 30 seconds.
- 2. Discard the flow-through and place the RB Column back in the 2 ml Collection Tube.
- 3. Prepare DNase I solution in a 1.5 ml microcentrifuge tube (RNase-free) as follows:

DNase I	5 μl (2 U/μl)
DNase I Reaction Buffer	45 µl
Total Volume	50 μl

- 4. Gently pipette DNase I solution to mix (DO NOT vortex) then add DNase I solution (50 μI) into the CENTER of the RB column matrix.
- 5. Incubate the column for 15 minutes at room temperature (20-30°C) then proceed with the RNA Wash step.



4. RNA Wash

Add **400** μ I of W1 Buffer to the RB Column then centrifuge at 14-16,000 x g for 1 minute. Discard the flow-through and place the RB Column back in the 2 ml Collection Tube. Add **600** μ I of Wash Buffer (make sure ethanol was added) into the RB Column. Centrifuge at 14-16,000 x g for 1 minute. Discard the flow-through then place the RB Column back in the 2 ml Collection Tube. Add **600** μ I of Wash Buffer (make sure ethanol was added) into the RB Column. Centrifuge at 14-16,000 x g for 1 minute. Discard the flow-through then place the RB Column back in the 2 ml Collection Tube. Centrifuge at 14-16,000 x g for 3 minutes to dry the column matrix.

RNA Elution

Place the **dried RB Column** in a clean 1.5 ml microcentrifuge tube (RNase-free). **Add 50 µl of RNase-free Water** into the **CENTER** of the column matrix. Let stand for at least 3 minutes to ensure the **RNase-free Water** is absorbed by the matrix. Centrifuge at 14-16,000 x g for 1 minute to elute the purified RNA.

Optional Step 2: DNA Digestion In Solution

1. Prepare DNase I reaction in a 1.5 ml microcentrifuge tube (RNase-free) as follows:

RNA in RNase-free Water	1-40 µl
DNase I	0.5 μl/μg RNA
DNase I Reaction Buffer	5 μl
RNase-free Water	Add to final volume = 50 μl
Total Volume	50 μl

- 2. Gently pipette the DNase I reaction solution to mix (DO NOT vortex) then incubate the microcentrifuge tube at 37°C for 15-30 minutes.
- 3. Stop the reaction by adding 1 μ I of 20 mM EGTA (pH=8.0) then incubate the microcentrifuge tube at 65°C for 10 minutes.

NOTE: DNase I Reaction Buffer may cause aberrant migration or smearing of RNA on gels. If analyzing RNA by gel electrophoresis, repurify the RNA sample by using the Geneaid™ RNA Cleanup Kit instead of stopping the reaction with EGTA.



Troubleshooting

Low Yield



Clogged Column.

Reduce the amount of starting material or separate it into multiple tubes. Centrifugation temperature must be between 20°C to 25°C. Yeast cells were not completely homogenized. Make sure lyticase or zymolase was added to Sorbitol Buffer immediately prior to use.

Residual Ethanol Contamination.

Following the wash step, dry the RB Column with additional centrifugation at 14-16,000 x g for 5 minutes.

RNA Degradation.

The harvested sample should be stabilized immediately prior to use. Disposable plasticware and automatic pipettes should be sterile (RNase-free) and used only for RNA procedures. Non-disposable glassware or plasticware should also be sterile (RNase-free).

Presto™ Mini RNA Yeast Kit Functional Test Data

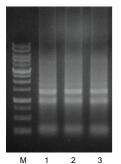


Figure 1. Total RNA was extracted using the Presto[™] Mini RNA Yeast Kit. *Saccharomyces cerevisiae* (5×10⁷) was harvested by centrifugation at 5,000 x g for 10 minutes. A 5 μl aliquot of purified RNA from a 50 μl eluate was analyzed by electrophoresis on a 0.8% agarose gel.

M = Geneaid 1 Kb DNA Ladder

Test	RNA Conc.	260/280	260/230	Yield
1	391.0 µg/ml	2.19	2.48	19.6 µg
2	389.9 µg/ml	2.19	2.51	19.5 µg
3	387.9 µg/ml	2.20	2.51	19.4 µg



Related DNA Extraction Products

RNA Purification		
Product	Package Size	Catalogue Number
Total RNA Mini Kit (Blood/Cultured Cell)	50/100/300 preps	RB050/100/300
Total RNA Mini Kit (Tissue)	50/100/300 preps	RT050/100/300
Total RNA Mini Kit (Plant)	50/100/300 preps	RP050/100/300
Presto™ Mini RNA Bacteria Kit	50/100/300 preps	RBB050/100/300
Presto™ Mini RNA Yeast Kit	50/100/300 preps	RBY050/100/300
miRNA Isolation Kit	50/100 preps	RMI050/100
GENEzol™ Reagent	50/100/200 rxns	GZR050/100/200
GENEzol™ TriRNA Bacteria Kit	50/100 rxns	GZB050/100
GENEzol™ TriRNA Pure Kit	50/100/200 preps	GZX050/100/200
TriRNA Pure Kit	50/100/200 preps	TRP050/100/200
RNA Cleanup Kit	50/100 preps	PR050/100
Virus DNA/RNA Purification		
Product	Package Size	Catalogue Number
Plant Virus RNA Kit	50/100 preps	PVR050/100
Viral Nucleic Acid Extraction Kit II	50/100/300 preps	VR050/100/300
Viral Nucleic Acid Extraction Kit III	50/100/300 preps	VI050/100/300
Genomic DNA Extraction		
Product	Package Size	Catalogue Number
Genomic DNA Mini Kit (Blood/Cultured Cell)	100/300 preps	GB100/300
Genomic DNA Maxi Kit (Blood/Cultured Cell)	10/25 preps	GDM010/25
Genomic DNA Mini Kit (Tissue)	50/100/300 preps	GT050/100/300
gSYNC™ DNA Extraction Kit	50/100/300 preps	GS050/100/300
Genomic DNA Mini Kit (Plant)	100 preps	GP100
Genomic DNA Maxi Kit (Plant)	10/25 preps	GPM10/25
GENEzol™ DNA Reagent Plant	100/200 rxns	GR100/200
Presto™ Mini gDNA Yeast Kit	100/300 preps	GBY100/300
Presto™ Mini gDNA Bacteria Kit	100/300 preps	GBB100/101/300/301
Geneius™ Micro DNA Extraction Kit	100/300 preps	GMB100/300
Presto™ Buccal Swab gDNA Extraction Kit	100/300 preps	GSK100/300
Presto™ 96 Well Blood Genomic DNA Extraction Kit	4/10 x 96 preps	96GBP04/10
Presto™ 96 Well Plant Genomic DNA Extraction Kit	4/10 x 96 preps	96GPP04/10
DNA RNA Purification		
Product	Package Size	Catalogue Number
Presto™ DNA/RNA Extraction Kit	50/100 preps	DR050/100
Presto™ DNA/RNA/Protein Extraction Kit	50/100 preps	DRP050/100

For additional product information please visit www.geneaid.com. Thank you!

Geneaid

